18-nor-Abietatrienes from the Cones of Larix kaempferi

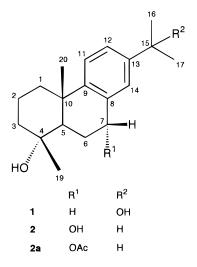
Hironori Ohtsu, Reiko Tanaka, and Shunyo Matsunaga*

Osaka University of Pharmaceutical Sciences, 4-20-1 Nasahara, Takatsuki, Osaka 569-1094, Japan

Received October 20, 1997

Two novel norditerpenediols, 18-*nor*-abieta-8,11,13-triene-4,15-diol (1) and 18-*nor*-abieta-8,-11,13-triene-4,7 α -diol (2), were isolated from the cones of *Larix kaempferi*, together with two known diterpenes, abieta-8,11,13-triene-15,18-diol and abieta-8,11,13-triene-7 α ,18-diol. The structures of 1 and 2 were determined on the basis of chemical and spectral evidence.

Previously, we reported the isolation of abieta-8,11,-13,15-tetraen-18-oic acid, 16-nor-15-oxodehydroabietic acid, and 12,15-dihydroxydehydroabietic acid from the leaves of Larix kaempferi (Lamb.) Carr. (Pinaceae), together with 12 known compounds, including (-)- α cadinol, oplopanone, larixol, two dehydroabietinols, and seven dehydroabitic acids including a pair of 9α , 13α epidioxy- and 9β , 13β -epidioxyabiet-8(14)-en-18-oic acids.¹ No detailed study, however, has so far been reported on extracts of the cones of Larix species. Careful fractionation of a CHCl₃ extract of fresh cones of L. kaempferi, by Si gel column chromatography and reversed-phase HPLC, furnished two new compounds, 1 and 2, and several known compounds. Two of the known compounds were identified as abieta-8.11.13triene-15,18-diol^{2,3} and abieta-8,11,13-triene-7α,18-diol⁴ by comparison of their IR, ¹H NMR, and ¹³C NMR, and EIMS data with those already published. We now report the characterization of 1 and 2.



Compound **1** was assigned the molecular formula $C_{19}H_{28}O_2$ by HREIMS. Its IR spectrum contained absorption bands, indicating a hydroxyl group and a benzene ring. The ¹H and ¹³C NMR spectra showed signals for a tertiary methyl group, a tertiary methyl group geminal to an oxygen function, two equivalent methyls of a hydroxylsopropyl group,⁵ and an aromatic ring characteristic of abieta-8,11,13-trienes. No signal was observed for a methine proton geminal to a hydroxyl

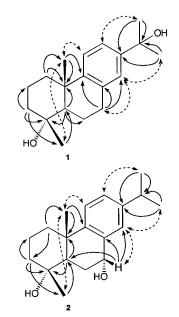


Figure 1. HMBC (solid arrow) and selected NOESY (dashed arrow) correlation of **1** and **2**.

group. Except for the presence of signals of a hydroxyisopropyl group and the absence of those for an isopropyl group, the above ¹H and ¹³C NMR spectral data were closely similar to those of the known compounds, 18nor-abieta-8,11,13-trien-4-ol6-8 and 19-nor-abieta-8,11,-13-trien-4-ol.^{7,8} The HMBC spectrum of **1** provided cross correlations (Figure 1), indicating that the two hydroxyl groups of 1 should be placed at C-4 and C-15. The chemical shift values of signals for Me-20, C-5, C-19, and C-20 were closely similar to those of 18-nor-abieta-8,11,13-trien-4-ol rather than those of 19-nor-abieta-8,-11.13-trien-4-ol. In the NOESY experiment (Figure 1). a significant correlation was observed between signals at δ 1.16 and 1.24, indicating Me-20 and Me-19 (geminal to a tertiary hydroxyl group) have a 1,3-diaxial relationship. Thus, the structure of **1** was proved to be 18-norabieta-8.11.13-triene-4.15-diol.

Compound **2** was also assigned the molecular formula $C_{19}H_{28}O_2$ by HREIMS. Its IR spectrum contained absorptions characteristic for hydroxyl and aromatic groups. The ¹H and ¹³C NMR spectra revealed signals for a tertiary methyl group, a tertiary methyl group geminal to a hydroxyl group, an isopropyl group, a benzene ring corresponding to the C-ring of **1**, and a hydroxymethine group. Acetylation of **2** afforded a

^{*} To whom correspondence should be addressed. Tel. and Fax: +81 726-90-1084. E-mail: matunaga@oysun01.oups.ac.jp.

Notes

hydroxymonoacetate (2a). The presence of an isopropyl group and a secondary hydroxyl group and the absence of a hydroxyisopropyl group suggested that 2 was a positional isomer of 1. This assumption was supported by the ${}^{1}H-{}^{1}H$ COSY, HMQC, and HMBC experiments. The HMBC data exhibited correlations as shown in Figure 1, indicating 2 to be either 18-nor- or 19-norabieta-8,11,13-triene-4,7 ξ -diol. The C-7 signal of **2** observed at $\delta_{\rm C}$ 68.1 was compatible with those reported for 7α -hydroxyabieta-8,11,13-trienes,^{9,10} which was shifted ca. 2 ppm to higher field than those of the known 7β -hydroxyabieta-8,11,13-trienes.^{9,11} Furthermore, the C-5 signal was shifted upfield 6.1 ppm from that of 1, whereas those of C-6, C-12, and C-14 were shifted 9.1, 4.6, and 3.1 ppm downfield, respectively, from those of **1** due to the addition of a 7α -hydroxyl group in the molecule. The NOESY data provided cross correlations as shown in Figure 1, indicating that the two hydroxyl groups of **2** have 4α - and 7α -configurations. Hence, compound 2 was proved to be 18-nor-abieta-8,11,13triene-4,7 α -diol.

Experimental Section

General Experimental Procedures. Melting points were determined on a Yanagimoto micromelting-point apparatus and were uncorrected. Optical rotations were measured using a JASCO DIP-1000 digital polarimeter. IR spectra were recorded as KBr disks using a Perkin-Elmer 1720X FTIR spectrophotometer. ¹H and ¹³C NMR spectra were obtained on a Varian XL-300 and INOVA 500 spectrometer with standard pulse sequences operating at 300 and 500 MHz and 74.5 and 125 MHz, respectively. CDCl₃ was used as solvent and TMS as internal standard. EIMS and HREIMS were recorded on a Hitachi 4000H double-focusing mass spectrometer (70 eV). Column chromatography was carried out over Si gel (70~230 mesh, Merck) and Cosmocil 75C₁₈-OPN (Nacarai Tesque), and MPLC was carried out with Cosmocil 40C₁₈-PREP (Nacarai Tesque). Preparative HPLC was carried out using a TOSOH system equipped with a CCPM-prep Pump, a SC-8020 system controller, and a TSK-GEL ODS-80Ts (21.5 \times 300 mm). Fractions obtained from column chromatography were monitored by TLC (Merck Si gel 60 HF₂₅₄). Preparative TLC was carried out on Merck Si gel PF₂₅₄ plates (0.5 mm).

Plant Material. The fresh cones of *L. kaempferi* were collected on the grounds of National Forest Tree Breeding Station, Komoro City, Nagano Prefecture, Japan, on July 30, 1995. Voucher specimens are deposited in the Herbarium of the Department of Medicinal Chemistry, Osaka University of Pharmaceutical Sciences, Takatsuki, Japan (voucher no. LK-957K).

Extraction and Isolation of Compounds. The fresh cones of *L. kaempferi* (1.45 kg) were chopped and extracted with CHCl₃ (4L \times 5). Evaporation of the extract in vacuo yielded a dark yellow residue (123 g), which was divided into 10 fractions by preliminary Si gel (2.0 kg) column chromatography using CHCl₃, CHCl₃–EtOAc (gradiently), EtOAc, and EtOAc–MeOH (1:1). Rechromatography of residue 9 collected from the EtOAc eluates (12.10 g) over a Si gel (200 g) column with CHCl₃–MeOH (100: 1) successively afforded residues **A** (484 mg) and **B** (859 mg) from the fraction nos. 29–37 and 38–44 (each fraction: 100 mL). Residue **A**

was purified by reversed-phase MPLC using Cosmosil $40C_{18}$ -PREP (ODS) column and 60% MeOH $-H_2O$ to give compound **2** (7 mg) as a crystalline solid. Residue **B** was subjected to ODS open column chromatography. Elution with 70% MeOH-H₂O afforded residues C (208 mg), **D** (42 mg), and **E** (68 mg) from the fraction nos. 1-12, 13-26, and 39-44 (each fraction: 30 mL). Residue C, purified by HPLC (ODS) using 65% MeOH $-H_2O$, furnished compound 1 (6 mg). Recrystallization of residue **D** from *n*-hexanes-EtOAc afforded abieta-8,-11,13-triene-15,18-diol, 30 mg, mp 100–101 °C, $[\alpha]^{26}$ $+2^{\circ}$ (c 0.50, EtOH). Residue E was purified by preparative TLC (n-hexane-EtOAc, 2:1) to afford abieta-8,11,13-triene-7α,18-diol, 26 mg (Me₂CO), mp 87–88 °C, $[\alpha]^{26}_{D}$ –1° (*c* 0.43, EtOH). Physical and spectral data of these latter two compounds were compatible with those already reported in the literature.²⁻⁴

18-nor-Abieta-8,11,13-triene-4,15-diol (1): colorless needles (*n*-hexanes–EtOAc); mp 140–141 °C; $[\alpha]^{26}$ -13° (c 0.32, CHCl₃); IR (KBr) ν_{max} 3279 (OH), 2970, 2930, 1610, 1498, and 1458 (aromatic ring), 1175, 986, 876 cm⁻¹; ¹H NMR (CDCl₃, 500 MHz) δ 7.24 (1H, dd, J = 8.0, 2.0 Hz, H-12), 7.22 (1H, d, J = 8.0 Hz, H-11), 7.17 (1H, d, J = 2.0 Hz, H-14), 2.93 (2H, m, H-7 α , H-7 β), 2.26 (1H, ddd, J = 13.0, 4.0, 2.8 Hz, H-1 β), 2.12 (1H, dddd, J = 13.0, 7.0, 1.5, 1.5 Hz, H-6 α), 1.88 (1H, dddd, J = 13.0, 3.5, 3.3, 1.5 Hz, H-3 β), 1.77 (1H, m, H-2 α), 1.70 (1H, m, H-6 β), 1.66 (1H, m, H-2 β), 1.60 (1H, m, H-5 α), 1.57 (6H, s, Me-16, Me-17), 1.44 (1H, ddd, J =13.0, 13.0, 4.0 Hz, H-1 α), 1.39 (1H, ddd, J = 13.0, 13.0, 4.5 Hz, H-3α), 1.24 (3H, s, Me-19), 1.16 (3H, s, Me-20); ¹³C NMR (CDCl₃, 125 MHz) δ 147.4 (s, C-9), 146.0 (s, C-13), 134.8 (s, C-8), 125.0 (d, C-14), 124.6 (d, C-11), 122.0 (d, C-12), 72.4 (s, C-4), 72.3 (s, C-15), 52.4 (d, C-5), 42.7 (t, C-3), 38.3 (s, C-10), 38.0 (t, C-1), 31.6 (q \times 2, C-16, C-17), 30.5 (t, C-7), 24.5 (q, C-20), 23.0 (q, C-19), 20.5 (t, C-2), 18.0 (t, C-6); EIMS (70 eV) m/z 288 [M]⁺ (29), 273 $[M - Me]^+$ (100), 255 $[273 - H_2O]^+$ (70), 227 (6), 197 (8), 185 (17), 157 (10), 131 (11), 43 (39); HREIMS m/z 288.2077 (calcd for C₁₉H₂₈O₂, 288.2088).

18-nor-Abieta-8,11,13-triene-4,7a-diol (2): colorless needles (*n*-hexane–EtOAc); mp 85–86 °C; $[\alpha]^{26}_{D}$ –15° (c 0.46, CHCl₃); IR (KBr) v_{max} 3371 (OH), 2965, 2931, 2867, 1568, 1497, and 1457 (aromatic ring), 1149, 983, 858 cm⁻¹; ¹H NMR (CDCl₃, 500 MHz) δ 7.19 (1H, d, J = 8.2 Hz, H-11), 7.16 (1H, d, J = 2.0 Hz, H-14), 7.12 (1H, dd, J = 8.2, 2.0 Hz, H-12), 4.81 (1H, dd, J = 4.0)1.7 Hz, H-7 β), 2.87 (1H, septet, J = 6.9 Hz, H-15), 2.23 (1H, br dd, J = 13.0, 1.7 Hz, H-6 α), 2.21 (1H, ddd, J =13.1, 3.8, 2.3 Hz, H-1 β), 1.94 (1H, br d, J = 13.0 Hz, H-5 α), 1.89 (1H, ddd, J = 13.0, 13.0, 4.0 Hz, H-6 β), 1.83 $(1H, dddd, J = 13.3, 3.3, 3.3, 1.2 Hz, H-3\beta), 1.73 (1H, J-1)$ m, H-2 α), 1.63 (1H, m, H-2 β), 1.37 (1H, ddd, J = 13.1, 13.1, 3.8 Hz, H-1 α), 1.32 (1H, ddd, J = 13.3, 13.3, 4.4 Hz, H-3 α), 1.24 (6H, d, J = 6.9 Hz, Me-16, Me-17), 1.18 (3H, s, Me-19), 1.08 (3H, s, Me-20); ¹³C NMR (CDCl₃, 125 MHz) & 146.5 (s, C-13), 146.4 (s, C-9), 135.9 (s, C-8), 128.1 (d, C-14), 126.6 (d, C-12), 124.7 (d, C-11), 71.8 (s, C-4), 68.1 (d, C-7), 46.3 (d, C-5), 42.6 (t, C-3), 38.7 (s, C-10), 37.7 (t, C-1), 33.5 (d, C-15), 27.1 (t, C-6), 24.0 (q, C-17), 23.9 (q, C-16), 23.5 (q, C-20), 22.5 (q, C-19), 20.5 (t, C-2); EIMS (70 eV) *m*/*z* 288 [M]⁺ (100), 255 [M – Me $(-H_2O)^+$ (41), 237 [255 $(-H_2O)^+$ (51), 195 (67), 185 (17),

143 (89), 109 (41), 85 (83), 43 (62); HREIMS m/z 288.2098 (calcd for C₁₉H₂₈O₂, 288.2088).

Compound 2 Acetate (2a). A mixture of compound **2** (2.4 mg) in dried pyridine–Ac₂O (1:1, 1 mL) was left at room temperature overnight. Workup as usual yielded a solid (2.8 mg) that was recrystallized to furnish monoacetate (2a), 2.0 mg (n-hexane-EtOAc), as colorless needles: mp 156–157 °C, $[\alpha]^{26}_{D}$ +4° (*c* 0.21, CHCl₃); IR (KBr) v_{max} 3553 (OH), 2932, 2867, 1726, and 1249 (OAc), 1497 and 1457 (aromatic ring) cm⁻¹; ¹H NMR $(CDCl_3, 500 \text{ MHz}) \delta 7.24 (1H, d, J = 8.2 \text{ Hz}, \text{H-}11), 7.17$ (1H, dd, J = 8.2, 2.0 Hz, H-12), 7.07 (1H, d, J = 2.0 Hz, H-14), 6.06 (1H, br d, J = 4.0 Hz, H-7 β), 2.86 (1H, septet, J = 7.0 Hz, H-15), 2.26 (1H, ddd, J = 13.0, 4.0, 2.3 Hz, H-1 β), 2.24 (1H, m, H-6 α), 2.07 (3H, s, OCOCH₃), 1.95 $(2H, m, H-5\alpha, H-6\beta), 1.90 (1H, ddd, J = 13.5, 3.0, 3.0)$ Hz, H-3 β), 1.79 (1H, m, H-2 α), 1.66 (1H, m, H-2 β), 1.48 $(1H, ddd, J = 13.0, 13.0, 4.0 Hz, H-1\alpha), 1.42$ (1H, ddd, J = 13.5, 13.5, 4.5 Hz, H-3 α), 1.22 (6H, d, J = 7.0 Hz, Me-16, Me-17), 1.22 (3H, s, Me-19), 1.11 (3H, s, Me-20); ¹³C NMR (CDCl₃, 125 MHz) δ 170.8 (s, OCOCH₃), 147.2 (s, C-9), 146.6 (s, C-13), 132.1 (s, C-8), 128.4 (d, C-14), 127.1 (d, C-12), 124.6 (d, C-11), 71.7 (s, C-4), 70.3 (d, C-7), 47.3 (d, C-5), 42.6 (t, C-3), 38.4 (s, C-10), 37.5 (t, C-1), 33.5 (d, C-15), 25.0 (t, C-6), 24.0 (q, C-17), 23.8 (q, C-16), 23.5 (q, C-20), 22.7 (q, C-19), 21.7 (q, OCOCH₃), 20.4 (t, C-2); EIMS (70 eV) m/z 330 [M]⁺ (1), 312 [M - H_2O]⁺ (22), 270 [M – AcOH]⁺ (94), 255 [270 – Me]⁺ (25), $237 [255 - H_2O]^+$ (33), 195 (30), 185 (21), 169 (34), 155 (24), 143 (57), 85 (34), 43 (100); HREIMS m/z 330.2187 (calcd for $C_{21}H_{30}O_3$, 330.2193).

Acknowledgment. This work was supported by National Osaka Forestry Bureau, the Ministry of Agriculture, Forestry and Fisheries, Japan, 1-8-75, Temmabashi, Kita-ku, Osaka 530-0042, Japan. The authors are grateful to Mr. Y. Nakatani and Mr. S. Itoh, Nagano Forest Tree Breeding Station, National Forest Tree Breeding Center, for the collection of plant materials. Our thanks are also due to Mr. K. Minoura and Mrs. M. Fujitake, Osaka University of Pharmaceutical Sciences, for NMR and MS measurements.

References and Notes

- Tanaka, R.; Ohtsu, H.; Matsunaga, S. Phytochemistry 1997, 46, 1051–1057.
- (2) Conner, A. H.; Rowe, J. W. Phytochemistry 1977, 16, 1777-1781.
- (3) Jurgens, A. L.; McChesney, J. D. Magn. Reson. Chem. 1990, 28, 181–184.
- (4) Ohmoto, T.; Kanatani (nèe Inagaki), K.; Yamaguchi, K. Chem. Pharm. Bull. 1987, 35, 229–234.
- (5) Pereda-Miranda, R.; Delgado, G.; Romo de Vivar, A. Phytochemistry 1986, 25, 1931–1933.
- (6) Seelye, R. N.; Watkins, W. B. *Tetrahedron* **1969**, *25*, 447–451.
- (7) Rowe, J. W.; Nagasampagi, B. A. Phytochemistry 1971, 10, 1647–1651.
- (8) Lee, C.-K.; Fang, J.-M.; Cheng, Y.-S. *Phytochemistry* **1995**, *39*, 391–394.
- (9) Del Corral, J. M. M.; Gordaliza, M.; Salinero, M. A.; San Feliciano, A. Magn. Reson. Chem. 1994, 32, 774-781.
- (10) Yano, S.; Furuno, T. Mokuzai Gakkaishi 1994, 40, 72-77.
- (11) Lee, C.-K.; Fang, J.-M.; Cheng, Y.-S. *Phytochemistry* **1994**, 35, 983–986.
- (12) Tanaka, O.; Mihashi, S.; Yanagisawa, I.; Nikaido, T.; Shibata, S. Tetrahedron 1972, 28, 4523–4537.

NP9704720